

Technical Bulletin

MetaPolyzyme Multilytic Enzyme Mix

MAC4L

Storage Temperature -20 °C

Product Description

Metagenomics is a rapidly expanding field of basic and applied research which looks at all DNA that has been isolated directly from given single samples (e.g. environmental samples, biological organisms).^{1,2} Metagenomics allows for the investigation of microbes that exist in extreme environments, and which have been historically difficult to isolate, culture, and study.³ Metagenomics has revealed the existence of novel microbial species.⁴ Applications of metagenomic studies include public health data analysis,^{5,6} discovery of novel proteins, enzymes and natural products,^{7,8} environmental studies,^{9,10} and agricultural investigations.^{11,12}

MetaPolyzyme is a mix of enzymes which have proven useful in metagenomic studies of microbiomes of extreme and unique environments.¹³⁻¹⁶ The product is intended for digestion of the microbes for evaluation by whole genome shotgun sequencing for metagenomics and metatranscriptomic approaches.

Microbes are difficult to disrupt because the cell walls may form capsules or resistant spores. DNA can be extracted by using lysing enzymes such as lyticase, chitinase, zymolase, and gluculase to induce partial spheroplast formation. Spheroplasts are subsequently lysed to release DNA. The original version of this enzyme blend was formulated with the intent to facilitate more effective lysis of spheroplasts (or protoplasts).¹⁶

This product was evaluated and developed in consultation and collaboration with the Association of Biomolecular Resource Facilities (ABRF) Metagenomics and Microbiome Research Group (MMRG; formerly the Metagenomics Research Group, MGRG).¹³⁻¹⁶ Several publications have cited use of this product.¹⁷⁻¹⁹

The enzymes in MetaPolyzyme are:

- Mutanolysin
- Achromopeptidase
- Lyticase
- Chitinase
- Lysostaphin
- Lysozyme

Mutanolysin (from *Streptomyces globisporus*)

Mutanolysin is a muralytic enzyme (muramidase) that cleaves the β -N-acetylmuramyl-(1 \rightarrow 4)-N-acetylglucosamine linkage of the bacterial cell wall peptidoglycan-polysaccharide, particularly the β (1 \rightarrow 4) bond in MurNAc-GlcNAc.²⁰ Mutanolysin particularly acts on many Gram-positive bacteria, where the enzyme's carboxy-terminal moieties participate in the recognition and binding of unique cell wall structures.

Achromopeptidase

Achromopeptidase (known also as β -lytic protease²¹) has potent bacteriolytic activity on many Gram-positive aerobic bacteria²² with high lytic activity, against bacterial strains with the A1a chemotype (e.g., *Aerococcus viridans*), and the A3a chemotype (e.g., *Staphylococcus epidermidis*) for cell wall peptidoglycan structures. The enzyme has been reported to have particular recognition for Gly-X sites in peptide sequences, and for Gly-Gly and D-Ala-X sites in peptidoglycans.²³

Lyticase (from *Arthrobacter luteus*)

Lyticase is useful in digestion of linear glucose polymers with $\beta(1\rightarrow3)$ linkages, of yeast glycan coats and for spheroplast formation, and of the cell wall of active yeast cells.

Chitinase (from *Streptomyces griseus*)

Chitinase degrades chitin by enzymatic hydrolysis to N-acetyl-D-glucosamine. Degradation occurs via two consecutive enzyme reactions:

- Chitodextrinase-chitinase, a poly(1,4- β -[2-acetamido-2-deoxy-D-glucoside])-glycanohydrolase, removes chitobiose units from chitin.
- N-acetylglucosaminidase-chitobiase cleaves the disaccharide to its monomer subunits, N-acetyl-D-glucosamine (NAGA).

Lysostaphin (from *Staphylococcus staphylolyticus*)

Lysostaphin is a lytic enzyme with activity against *Staphylococcus* species, including *S. aureus*. Lysostaphin has hexosaminidase, amidase, and endopeptidase activities. It cleaves polyglycine crosslinks in the cellular wall of *Staphylococcus* species, which leads to cell lysis.^{24,25}

Lysozyme (from chicken egg white)

Lysozyme hydrolyzes $\beta(1\rightarrow4)$ linkages between N-acetylmuraminic acid and N-acetyl-D-glucosamine residues in peptidoglycan, and between N-acetyl-D-glucosamine residues in chitodextrin. Lysozyme lyses the peptidoglycan cell wall of Gram-positive bacteria.²⁶

Precautions and Disclaimer

For R&D use only. Not for drug, household, or other uses. Please consult the Safety Data Sheet for information regarding hazards and safe handling practices.

Preparation Instructions

Dissolve the contents of MetaPolyzyme by adding 1 mL of PBS, pH 7.5, to make a 5 mg/mL solution. There should be no EDTA, Calcium or Magnesium present in solution.

Storage/Stability

This product ships at cooler temperature conditions ('wet ice'). Long-term storage at $-20\text{ }^{\circ}\text{C}$ is recommended. Reconstituted solutions of MetaPolyzyme may be stored at $-20\text{ }^{\circ}\text{C}$, but long-term solution stability has not been examined.

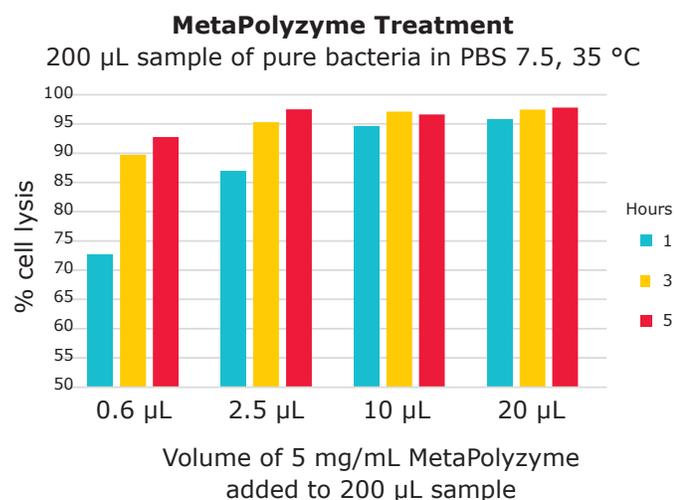
Procedure

Because of the great diversity of samples for metagenomics studies, it will be necessary for each researcher to work out particular conditions for optimal sample preparation and treatment. The following items should be noted before sample preparation:

- PBS buffer used for MetaPolyzyme and sample preparation should be at pH 7.5 and must not have EDTA, calcium or magnesium present in the solution.
- Sample with MetaPolyzyme must not be combined with Proteinase K.
- Sample with MetaPolyzyme must not be combined with lysis buffer or detergent.
- Sample suspected of containing interfering conditions mentioned above may be washed in 2X PBS before resuspending in PBS and adding MetaPolyzyme.
- MetaPolyzyme is also compatible with additional Lysozyme.
- It is recommended to also prepare 2 controls of PBS solution: one with MetaPolyzyme and one without MetaPolyzyme, as needed.
- The procedure below describes sample preparation in 2 mL microcentrifuge tubes. This procedure can be used as a guide to scale appropriately (ex: 96 well plate, swab samples).

1. Add 1 mL of PBS, pH 7.5 (without EDTA, calcium, magnesium) to MetaPolyzyme to create 5 mg/mL solution, set aside until ready for step 6.
Recommended: Aliquot MetaPolyzyme solution into desired aliquots for future use (example: 10 vials of 100 μ L) Aliquots can be stored in $-20\text{ }^{\circ}\text{C}$, but long-term stability of solution has not been examined. If solution will sit for more than 4 hours, add sodium azide to 0.02%.
2. Thoroughly suspend sample in PBS, pH 7.5.
3. Add 200 μ L of sample in PBS to 2 mL polypropylene microcentrifuge tube.
4. Optional pellet wash: To sample tube, add 1 mL of PBS, pH 7.5 Vortex, centrifuge and remove supernatant. Repeat step 4 two more times.
5. Resuspend pelleted sample in 150 μ L of PBS, pH 7.5 and vortex thoroughly.
6. Add *5-20 μ L of MetaPolyzyme to sample solution.
7. Incubate at $35\text{ }^{\circ}\text{C}$ for 2-24 hours.
8. Follow your standard DNA extraction protocol.

* Application data collected by the ABRF:



Each 200 μ L of pure bacteria sample was treated with 0.6 μ L, 2.5 μ L, 10 μ L or 20 μ L. The MetaPolyzyme + sample was incubated for 1, 3, and 5 hours at $35\text{ }^{\circ}\text{C}$. An OD reading of each sample per timepoint was recorded, as well as a sample with no MetaPolyzyme and a control of PBS only. A decrease in OD indicates an increase in cell lysis, so data is expressed as a % cell lysis, compared to the sample with no MetaPolyzyme (which would have 0% lysis). The treatment data shows that concentration and incubation time affect the % cell lysis of pure bacteria samples. Therefore, it is recommended that each user works out specific conditions for optimal sample preparation and treatment.

Notice

We provide information and advice to our customers on application technologies and regulatory matters to the best of our knowledge and ability, but without obligation or liability. Existing laws and regulations are to be observed in all cases by our customers. This also applies in respect to any rights of third parties. Our information and advice do not relieve our customers of their own responsibility for checking the suitability of our products for the envisaged purpose.

The information in this document is subject to change without notice and should not be construed as a commitment by the manufacturing or selling entity, or an affiliate. We assume no responsibility for any errors that may appear in this document.

Contact Information

For the location of the office nearest you, go to SigmaAldrich.com/offices.

Technical Assistance

Visit the tech service page on our web site at SigmaAldrich.com/techservice.

Standard Warranty

The applicable warranty for the products listed in this publication may be found at SigmaAldrich.com/terms.

References

1. Gilbert, J.A., and Dupont, C.L., *Ann. Rev. Marine Sci.*, **3**, 347-371 (2011).
2. Kang, H.S., and Brady, S.F., *J. Am. Chem. Soc.*, **136(52)**, 18111-18119 (2014).
3. Ufarté, L. et al., *Biotechnol. Adv.*, **33(8)**, 1845-1854 (2015).
4. Davison, M. et al., *Photosynth. Res.*, **126(1)**, 135-146 (2015).
5. Afshinnekoo, E. et al., *Cell Syst.*, **1(1)**, 72-87 (2015).
6. The MetaSUB International Consortium, *Microbiome*, **4**, 24 (2016). *Microbiome*, **4**, 45 (2016).
7. Trinidad, M. et al., *Front Microbiol.*, **6**, 890 (2015).
8. Coughlan, L.M. et al., *Front Microbiol.*, **6**, 672 (2015).
9. Palomo, A. et al., *ISME J.*, **10(11)**, 2569-2581 (2016).
10. Pold, G. et al., *Appl. Environ. Microbiol.*, **82(22)**, 6518-6530 (2016).
11. Mitra, N. et al., *J. Gen. Virol.*, **97(8)**, 1771-1784 (2016).

12. Theuns, S. *et al.*, *Infect. Genet. Evol.*, **43**, 135-145 (2016).
13. Baldwin, D.A. *et al.*, "Life at the Extreme", ABRF Metagenomics Research Group Poster 2015, presented at the ABRF 2015 Conference, St. Louis, MO, USA, March 28-31, 2015.
14. Baldwin, D.A. *et al.*, "Implementing New Standards in Metagenomics and the Extreme Microbiome Project", ABRF Metagenomics Research Group Poster 2016, presented at the ABRF 2016 Conference, Fort Lauderdale, FL, USA, February 20-23, 2016.
15. McIntyre, A. *et al.*, "Life at the Extreme: The ABRF Metagenomics Research Group", ABRF Metagenomics Research Group Poster 2017, presented at the ABRF 2017 Conference, San Diego, CA, USA, March 25-28, 2017.
16. Tighe, S. *et al.*, *J. Biomol. Tech.*, **28(1)**, 31-39 (2017).
17. Al-Hebeshi, N.N. *et al.*, *J. Oral Microbiol.*, **11(1)**, 1557986 (2018).
18. Zaikova, E. *et al.*, *Front. Ecol. Evol.*, 7, doi.org/10.3389/fevo.2019.00001 (2019).
19. Hamner, S. *et al.*, *Int. J. Environ. Res. Public Health*, **16(7)**, pii: E1097 (2019).
20. Gründling, A., and Schneewind, O., *J. Bacteriol.*, **188(7)**, 2463-2472 (2006).
21. Li, S.L. *et al.*, *J. Bacteriol.*, **172(11)**, 6506-6511 (1990).
22. Ezaki, T., and Suzuki, S., *J. Clin. Microbiol.*, **16(5)**, 844-846 (1982).
23. Li, S. *et al.*, *J. Biochem.*, **124(2)**, 332-339 (1998).
24. Browder, H.P. *et al.*, *Biochem. Biophys. Res. Commun.*, **19**, 383-389 (1965).
25. Robinson, J.M. *et al.*, *J. Bacteriol.*, **137(3)**, 1158 (<https://abrf.org/sites/default/files/temp/RGs/MG1164>) (1979).
26. Vocolo, D.J. *et al.*, *Nature*, **412(6849)**, 835-838 (2001).

Merck and Sigma-Aldrich are trademarks of Merck KGaA, Darmstadt, Germany or its affiliates. All other trademarks are the property of their respective owners. Detailed information on trademarks is available via publicly accessible resources.
© 2021 Merck KGaA, Darmstadt, Germany and/or its affiliates. All Rights Reserved.

The life science business of Merck operates
as MilliporeSigma in the U.S. and Canada.

The Merck logo is displayed in a bold, red, sans-serif font.