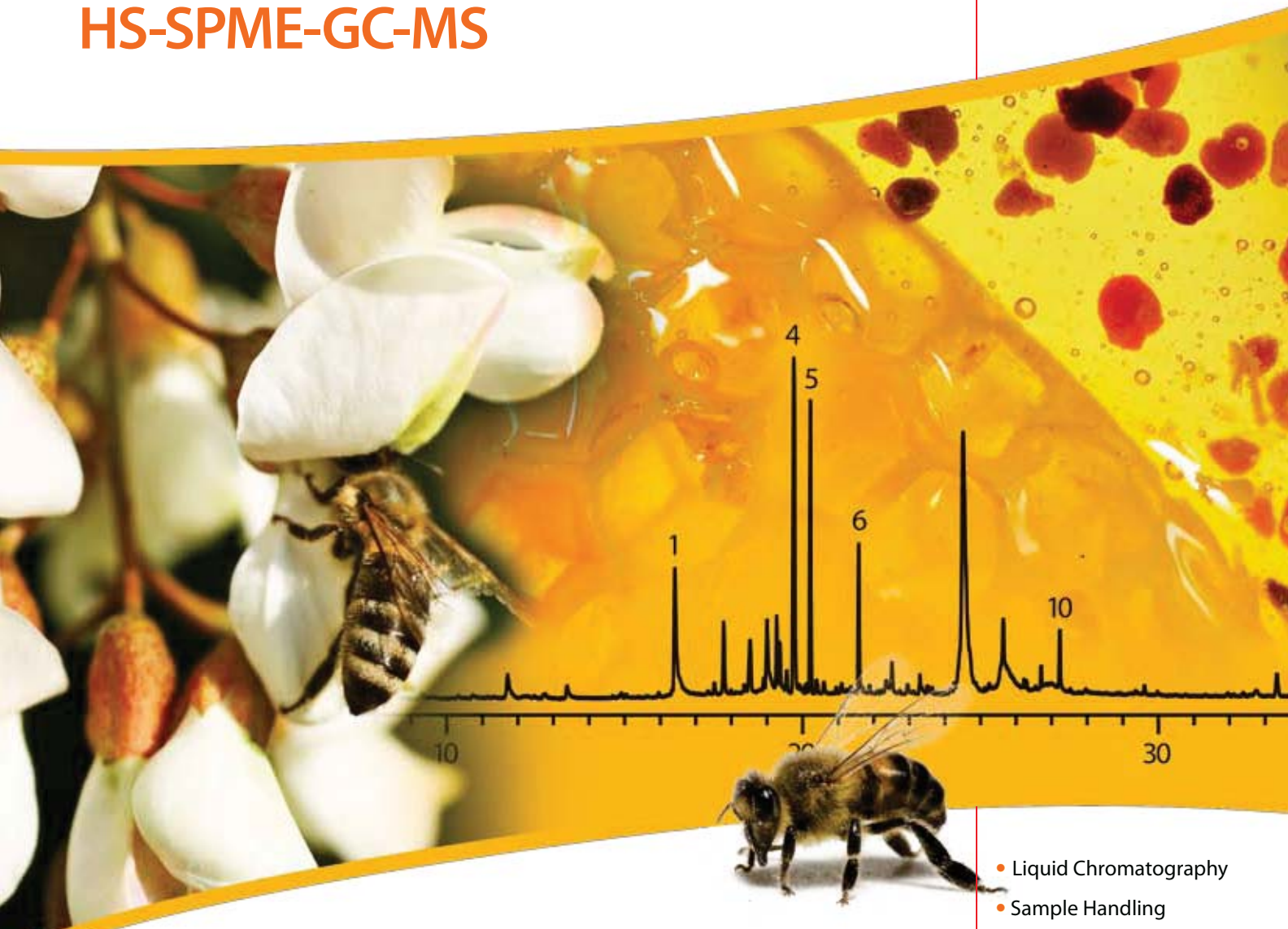


Reporter

Volume 28.1



Volatile Profiles of Honeys Using HS-SPME-GC-MS



The applicability of GC for determining botanical origin of honey

- Liquid Chromatography
- Sample Handling
- Gas Chromatography
- Standards
- Accessories
- Chiral Chromatography

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Don Hobbs

Director of Marketing

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Hope the new year is going well for you.

Regards,

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Volatile Profiles of Honey Using HS-SPME-GC-MS

Contributed Article

The following was generated with the assistance of an outside source using Sigma-Aldrich products. Technical content was generated and provided by:

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Introduction

The assessment of the authenticity of honey is a subject of primary importance both for consumers and industries. Usually, the determination of the botanical origin of honey is carried out by melissopalynological analysis, based on the identification of pollen by microscopic examination (1). However, data interpretation is very difficult and does not always lead to correct identification. The characterization of the volatile profile of honey has proven to be a reliable alternative to pollen analysis for the assessment of its botanical origin (2,3). In fact, the volatile profile represents a chemical fingerprint of honey, since both the nature and the amount of volatile compounds are characteristics of the floral source.

Solid phase microextraction (SPME) (4) is a rapid, solvent-less, and easy-to-use extraction technique widely applied in the determination of volatile compounds in several kinds of food. In this work we evaluate the applicability of headspace SPME (HS-SPME) coupled with gas chromatography-mass spectrometry (GC-MS) for the characterization of the volatile fraction of some honey samples.

Materials and Methods

Four samples of unifloral honeys (milk thistle, citrus, eucalyptus, and acacia) and a sample of a multifloral honey were analyzed. Dynamic headspace (DHS) extraction was performed following the procedure described in a previous work (2). A brief description of this DHS process can be found in Figure 1. HS-SPME was carried out per the procedure described in Figure 2. GC-MS conditions are listed in both Figures 1 and 2.

Compounds were identified by comparing the mass spectra obtained with those stored in the National Institute of Standards and Technology (NIST) library. In addition, retention indices (RIs) were calculated for each

(continued on page 4)

Table 1. Most Abundant Compounds Identified in Honey Headspace

Peak ID	Compound	Peak ID	Compound
1.	Nonanal	7.	Phenylacetaldehyde
2.	Furfural	8.	Nonanol
3.	Decanal	9.	3-Phenyl furan
4.	Benzaldehyde	10.	Phenylethyl alcohol
5.	α -Linalool	11.	Nonanoic acid
6.	Hotrienol		

Figure 1. Milk Thistle Honey Volatiles Using DHS

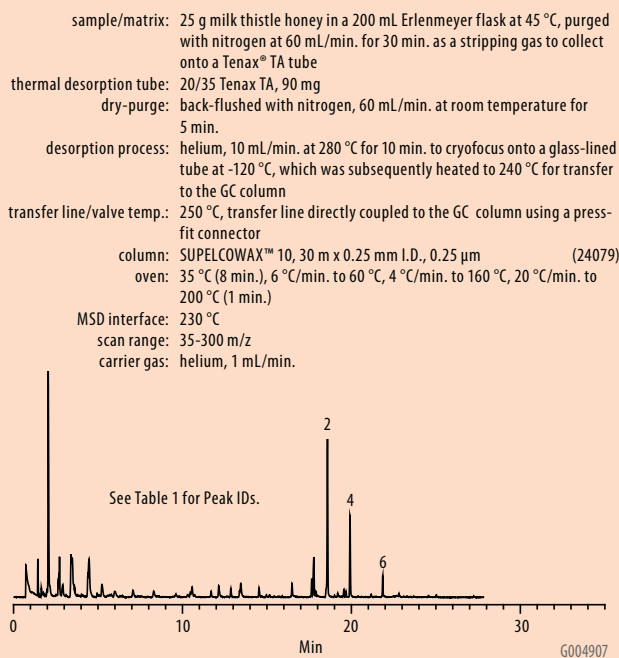


Figure 2. Milk Thistle Honey Volatiles Using SPME

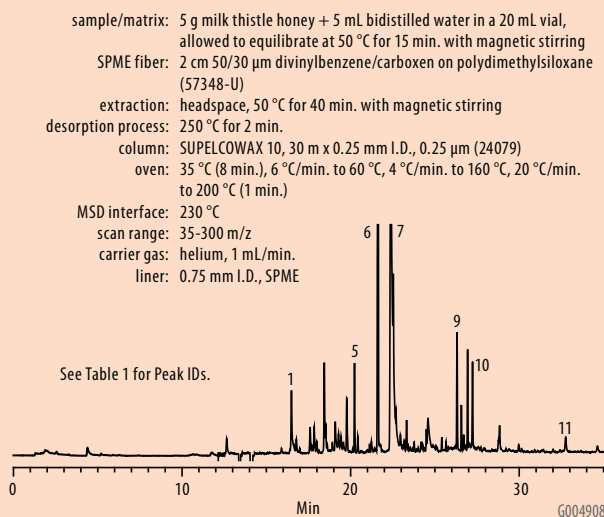
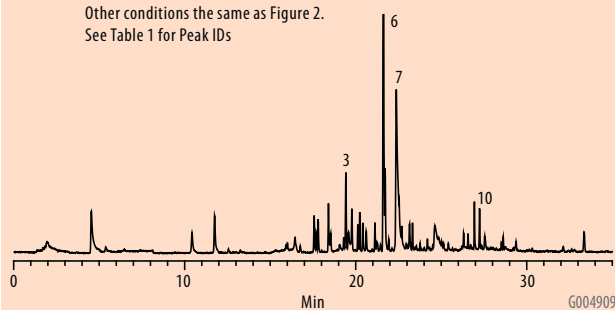


Figure 3. Citrus Honey Volatiles Using SPME

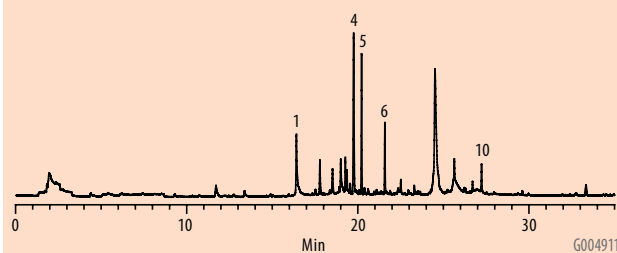
sample/matrix: 5 g citrus honey + 5 mL bidistilled water in a 20 mL vial, allowed to equilibrate at 50 °C for 15 min. with magnetic stirring

Other conditions the same as Figure 2.
See Table 1 for Peak IDs

**Figure 5. Acacia Honey Volatiles Using SPME**

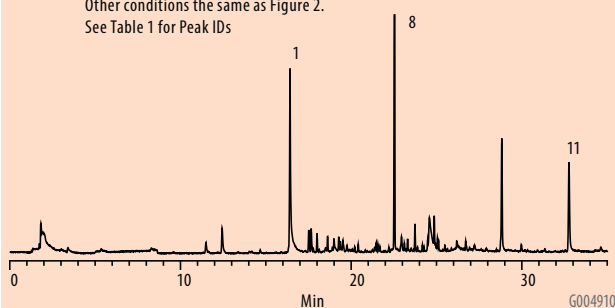
sample/matrix: 5 g acacia honey + 5 mL bidistilled water in a 20 mL vial, allowed to equilibrate at 50 °C for 15 min. with magnetic stirring

Other conditions the same as Figure 2.
See Table 1 for Peak IDs

**Figure 4. Eucalyptus Honey Volatiles Using SPME**

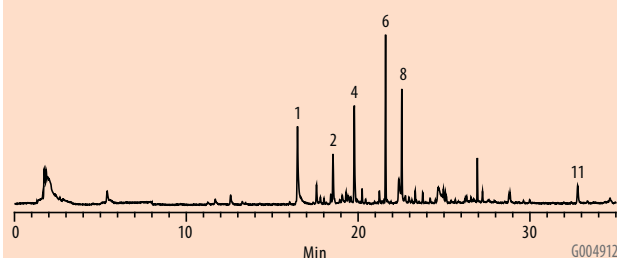
sample/matrix: 5 g eucalyptus honey + 5 mL bidistilled water in a 20 mL vial, allowed to equilibrate at 50 °C for 15 min. with magnetic stirring

Other conditions the same as Figure 2.
See Table 1 for Peak IDs

**Figure 6. Multifloral Honey Volatiles Using SPME**

sample/matrix: 5 g multifloral honey + 5 mL bidistilled water in a 20 mL vial, allowed to equilibrate at 50 °C for 15 min. with magnetic stirring

Other conditions the same as Figure 2.
See Table 1 for Peak IDs



Chromatograms for Figures 1-6 courtesy of Dr. Federica Bianchi and Prof. Marilena Musci (Univ. of Parma, Italy)

(continued from page 3)

peak and compared with literature data (5). Table 1 lists the most abundant compounds identified during this work. The chromatograms in Figures 1-6 are shown on the same abundance scale for ease of visual comparison. In order to evaluate quantitative differences in the aromatic profile of the samples investigated, gas chromatographic peak areas were calculated as Total Ion Current (TIC).

Results and Discussion

Preliminary experiments were carried out in order to compare the performances of the DHS technique, used in a previous study dealing with the characterization of the volatile profile of honey (2), with the SPME technique. Chromatographic profiles for a milk thistle honey obtained by DHS and SPME are shown in Figure 1 and Figure 2, respectively. As can be seen, the SPME profile not only shows a greater number of peaks with respect to that obtained by DHS, but also presents higher signal intensities. Less volatile compounds were better extracted using SPME, owing to its coating properties, whereas the more volatile compounds were better extracted by

DHS. Due to the greater number of peaks observed, SPME was chosen as the extraction technique for the remainder of our study.

Figures 2-6 show the chromatographic profile of five honey samples obtained by the SPME technique. From a qualitative point of view, the chromatographic profiles of honey samples of different botanical origins were quite similar, as some of the same volatile compounds were detected in different samples. So, it was not possible to define specific volatile compounds as markers characteristic for a defined botanical origin.

However, significant differences were observed from a quantitative point of view, since different amounts of the volatile compounds were observed depending upon the floral source. For example, the milk thistle honey (Figure 2) contained a high content of hotrienol (35%), phenylacetaldehyde (16%), and 3-phenyl furan (5%), the last compound being detected only in milk thistle honey. Citrus honey (Figure 3) was characterized by a high content of hotrienol (30%) and phenylacetaldehyde (10%). Alternatively, neither hotrienol nor phenylacetaldehyde were observed in eucalyptus honey (Figure 4), which was characterized by

high amounts of nonanol (32%), nonanal (12%), and nonanoic acid (7%). Acacia honey (Figure 5) was characterized by high amounts of benzaldehyde (18%), α -linalool (11%), nonanal (6%), and hotrienol (6%). The chromatographic profile of multifloral honey (Figure 6) presents a high number of compounds, the most abundant being hotrienol (14%), benzaldehyde (10%), nonanol (10%), nonanal (8%), and furfural (5%).

Conclusion

Considering both the nature and the relative amount of the volatile compounds extracted by SPME, it was possible to obtain and distinguish the chemical fingerprints of different types of honey. Therefore, it is possible that SPME may prove to be a useful tool for determining the botanical origin of honeys to achieve authenticity assessment.

References

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4. J. Pawliszyn, *Solid Phase Microextraction*; Wiley-VCH: New York, 1997.
5. Bianchi, F., Careri, M., Mangia, A. & Musci, M. (2007) *Journal of Separation Science*, 30, 563-572.

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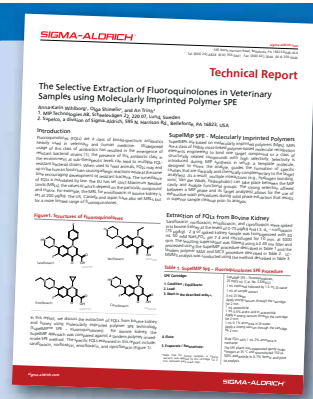
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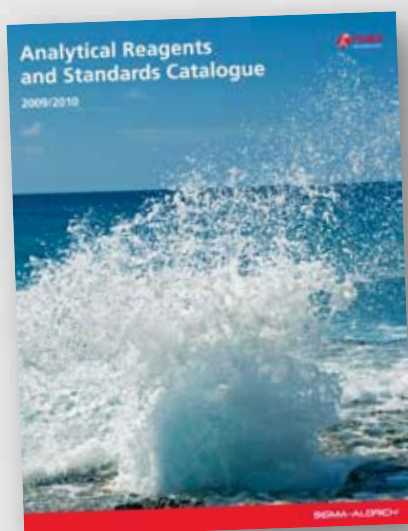
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HPLC & LC-MS: Techniques to Improve Speed, Resolution and Sensitivity without Investing in Capital Equipment

Chromatography column suppliers have responded to analysts’ need for higher throughput, better sensitivity, greater resolution and longer lifetimes through an array of innovations in particles, surface chemistries and column formats. While choice is good, its scope can often cloud the important underlying fundamentals: What do all these techniques mean? Why do they work? When would you choose one over the other? What about instrument compatibility? What role does mobile phase play? This talk will attempt to clarify the situation, demystifying the recent approaches the column industry has taken, and putting the innovations in historical and physicochemical context. Intended as an instructional talk, the scientific principles behind today’s innovative technology, such as Fused-Core™ particles, will be described and demonstrated using industry-specific applications.

Special Topics in Chromatography and Sample Prep

In this segment we will use special guest speakers to introduce current chromatography and sample prep topics. Title and speaker will be announced closer to the time of the event. Please consult the web site for up-to-date information.

Sample Prep for Chromatography: Sorbents, Devices and Techniques to Improve Sensitivity, Specificity and Throughput

Sample prep can be a significant bottleneck, especially when dealing with difficult or complex matrixes like biological fluids, tissues, food and agricultural products, grease and oils. This talk deals with unique combinations of innovative sorbents, devices and methodologies that address specific sample prep challenges. Topics include HybridSPE™-PPT technology for the low-level MS detection of pharmaceuticals and other small molecules in serum, solid phase microextraction (SPME), which has now been applied to bioanalysis, molecular imprinted polymers (SupelMIP™) and others. We will describe the scientific principles behind these technologies and demonstrate them using market-specific applications derived from the registrants’ areas of interest.

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Ascentis Express Peptide ES-C18 Expands the Fused-Core Particle Platform into Bioseparations

Wayne K. Way and William Campbell
wayne.way@sial.com

Ascentis Express Peptide ES-C18 columns were specifically engineered to separate higher molecular weight compounds such as peptides and small proteins. These columns contain advanced Fused-Core particles that have bigger pores (160 Å versus 90 Å in standard Ascentis Express), which greatly expands the application range for Ascentis Express columns.

Key Applications for Ascentis Express Peptide ES-C18:

- Pharmaceutical/therapeutic peptides
- Peptide mapping
- Natural and synthetic peptide analysis
- Oligonucleotide analysis

Key Advantages:

- Higher peak capacity providing greater resolution
- Amenable to higher flow rates for faster analysis
- Exceptional ruggedness providing long column lifetime

Ascentis Express Peptide ES-C18 columns utilize a steric-protected C18 bonded-phase with extremely high resistance to acid-catalyzed hydrolysis of the siloxane bond that attaches the C18 chain to the surface. Thus, the combination of low pH and elevated temperature operation of the column is well tolerated. Peptide separations are efficiently conducted using low pH mobile phase modifiers, often at 0.01-0.1% concentration, most popularly employing trifluoroacetic acid (TFA), and the related perfluorocarboxylic acids, pentafluoropropionic acid (PFPA) and heptafluorobutyric acid (HFBA). These acids exhibit desirable low UV transparency, volatility, and peptide ion-pairing properties. Additional opportunities for low pH operation include the normal short chain carboxylic acids, formic acid and acetic acid, as well as mineral acids, such as phosphoric acid (0.001-0.02 M).

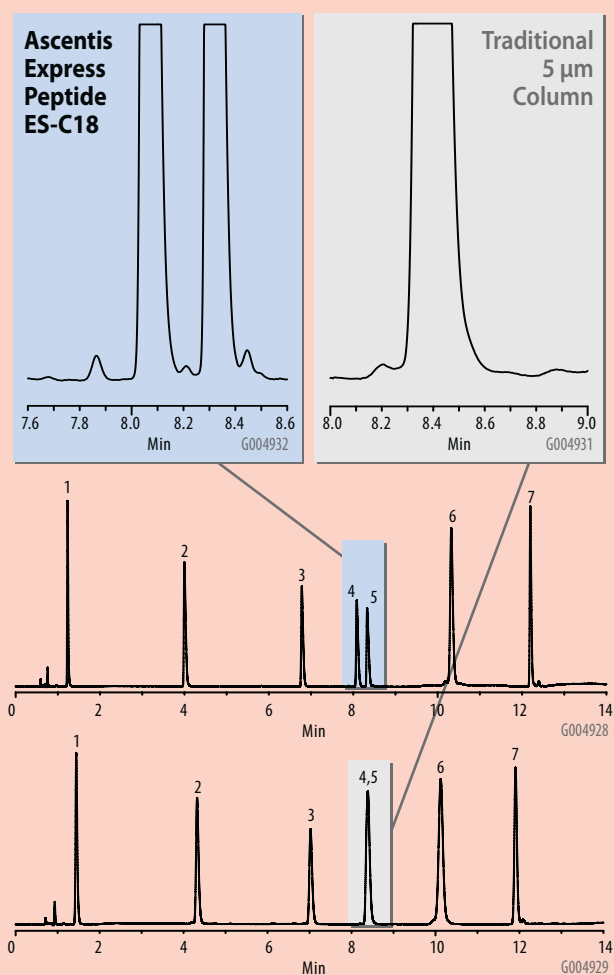
Shown in Figure 1 is the chromatographic separation of a peptide mix. The peptide mix contains a range of peptides in terms of molecular weight, basicity, and hydrophobicity. Excellent peak shape and peak width are achieved with a standard acetonitrile gradient and 0.1% TFA modifier. The resolution of small baseline impurities are shown in the inset, demonstrating the resolving power of the Ascentis Express Peptide ES-C18 column versus a traditional 5 µm column.

Figure 1. Comparison of Peptide Test Mix with Ascentis Express Peptide ES-C18 and Traditional Column

column: Ascentis Express Peptide ES-C18, 10 cm x 4.6 mm I.D. (53324-U)
mobile phase A: 10% acetonitrile / 90% water / 0.1% trifluoroacetic acid
mobile phase B: 75% acetonitrile / 25% water / 0.1% trifluoroacetic acid
gradient: 0% to 50% B in 15 min
flow rate: 1.5 mL/min.
det: UV at 220 nm
temp: 30 °C
injection: 5 µL

Peptide Test Mix

- | | | | |
|-------------------|-----------|-------------------|-------------|
| 1. Gly-Tyr | MW = 252 | 5. Leu-Enkephalin | MW = 555.62 |
| 2. Val-Tyr-Val | MW = 379 | 6. Ribonuclease | MW = 13,700 |
| 3. Met Enkephalin | MW = 574 | 7. Bovine Insulin | MW = 5733 |
| 4. Angiotensin II | MW = 1032 | | |



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3.0	53308-U	53311-U	53312-U	53313-U	53314-U
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Liquid Chromatography

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Flexible Method Development Using a New pH Stable HPLC Column, Kromasil Eternity C18

Jared Benedict, Marketing and Sales Manager, Kromasil
jared.benedict@akzonobel.com

Introduction

The separation of new generation pharmaceutical compounds has increasingly become more challenging. There is a need for stationary phases that can withstand a wide pH range, from very acidic to very basic conditions, while maintaining peak shape, symmetry and lifetime. Utilizing pH as a method development tool has now allowed more flexibility in altering retention times of ionizable compounds. More basic compounds are being synthesized during the discovery of pharmaceuticals and thus there is a great need for robust stationary phases when running at high pH. This article shows how Kromasil Eternity provides solutions for the separation of ionizable compounds using extreme pH conditions, from pH 1 to pH 12.

Results

In this study, we show the retention time of amines and acids can be altered by changing the pH, utilizing a single column (Figure 1). Running under basic (pH 11.0) conditions will destroy classic C18 phases very quickly, thus limiting the range of pH that can be utilized. However, when running

basic pH (11.0) conditions on a Kromasil Eternity C18, procaine, an amine, is strongly retained whereas the acidic nitrobenzoic acid is weakly retained. The reverse is true under acidic conditions. Therefore, not only can you utilize the solvent strength and temperature as a method development tool but you can now also utilize nearly the full pH range. The most striking conclusion from Figure 1 is that these extreme pH conditions can be run on a single column. Kromasil Eternity can be used at an extended pH range while maintaining a very long lifetime.

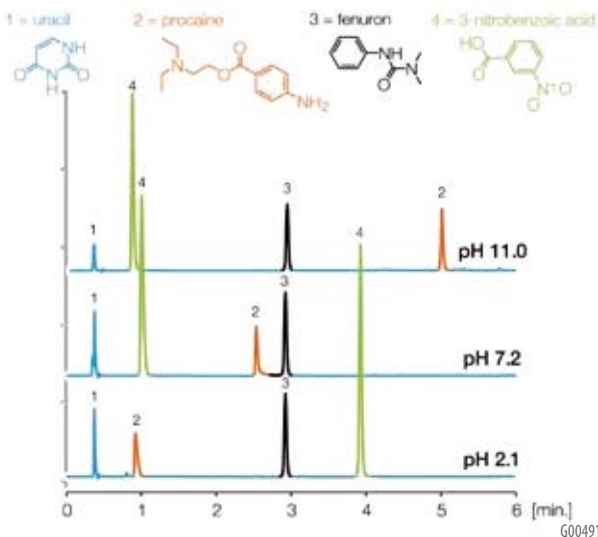
In addition to utilizing the flexibility in pH in analytical method development one can also employ the robustness of these columns for semi-preparative HPLC. When doing high throughput purification of pharmaceutical compounds one needs a column that can run under extreme pH conditions to maximize the separation factor and thus the loadability. Kromasil Eternity C18 provides a solution for high loadability and flexibility under extreme pH conditions using a single column which also maintains a long lifetime that gives the maximum total economy for your purification.

Conclusion

Kromasil Eternity provides flexible method development and long column lifetime through the use of unique modified silica. Kromasil Eternity C18 is available as a 2.5 μm particle size for UHPLC or HPLC applications. The small particle size allows for fast runs that maximize efficiency and minimize solvent consumption. Additionally, a 5 μm particle size is available for analytical to semi-preparative HPLC up to 30 mm I.D. columns. The loadability and lifetime of the semi-preparative columns are excellent.

Figure 1. Separation of Ionizable Compounds at Three Different pH Conditions

column: Kromasil Eternity C18, 5 cm x 4.6 mm, 2.5 μm
mobile phase: acetonitrile:20 mM sodium phosphate, pH 2.1, 7.2, and 11.0, respectively
gradient: 0-0.5 min: 10% acetonitrile, 5.5 min: 50% acetonitrile
flow rate: 1.5 mL / min
temp.: 25°C
det.: UV 254 nm
sample: 1. Uracil, 2. Procaine, 3. Fenuron, 4. 3-nitrobenzoic acid



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2.5	4.6	5	K08670896
2.5	4.6	10	K08670897
5	2.1	5	K08670898
5	2.1	15	K08670899
5	4.6	5	K08670900
5	4.6	10	K08670901
5	4.6	15	K08670902
5	4.6	25	K08670903

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Accelerated Bioanalytical LC-MS-MS Using Ascentis Express Fused-Core HPLC Columns

Wayne K. Way and Carmen T. Santasania

wayne.way@sial.com

Fast bioanalysis requires the use of short columns packed with high efficiency particles. The use of HPLC columns packed with Fused-Core particles and sub-2 μm porous particles have led the way in these demanding analyses. They significantly enhance resolution and speed by producing either higher efficiency for the same column length or equivalent efficiency with a shorter column length. The use of ultra-high performance liquid chromatography (UHPLC) comes at a price of much higher pressure when sub-2 μm particles are employed. As an attractive alternative, the revolutionary Ascentis Express 2.7 μm Fused-Core silica particle has quickly become accepted because it is equivalent in performance to particles in the sub-2 μm range. With a very narrow particle size distribution, Ascentis Express columns employ conventional 2 μm frits and operate ruggedly at much lower pressures that are within the operating limits of conventional HPLC instruments.

The goal of this work was to demonstrate the ease of converting traditional multi-minute assays on a 50 mm Discovery® C18 column to one-minute assays using 20 mm Ascentis Express columns on an HPLC configured for LC-MS-MS bioanalysis (Agilent 1100/ABI 3200 Q Trap MS-MS). The assay integrity and quality had to be maintained in the conversion. In addition, samples were extracted from plasma based on a published method (1) and analyzed by LC-MS-MS. These extraction conditions are shown in Table 1.

Table 2 shows the compounds, their MS-MS transitions, columns and chromatographic conditions used. The slopes of each gradient were adjusted to accommodate the 20 and 50 mm column lengths. Flow rates differ because the Discovery C18 5 μm column is typically used at 0.5 mL/min in these analyses while a 1 mL/min flow rate would be more likely used with the Ascentis Express to take advantage of the higher efficiency that can be obtained at this higher flow rate.

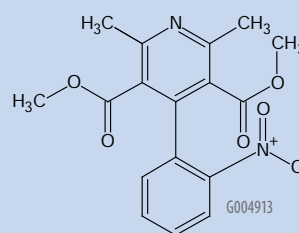
Figure 1a shows the compound mix separated in less than 0.5 minutes on the Ascentis Express HPLC column. Figure 1b shows the separation taking about 2 minutes. Excellent resolution is seen in both bases. Table 3 shows the peak width at half height measurement for both compounds. As expected the Ascentis Express phase shows much narrower peak width, indicating better efficiency for this column, even at the flow rate of 1 mL/min in a 2.1 mm column.

Table 1. Sample Preparation Procedure (1)

1. Prepare 5000 ng/mL sample in rat plasma by taking 5 μL of a mix carbamazepine and dehydronifedipine (1mg/mL each in methanol) and spike 1 mL of rat plasma.
2. Vortex 1 min, take 25 μL of plasma and 200 μL of acetonitrile to crash proteins.
3. Centrifuge at 10,000 rpm for 1 minute. Remove 100 μL supernatant and add 200 μL of water. Vortex 1 minute.
4. Inject 2 μL .

Table 2. Experimental Conditions

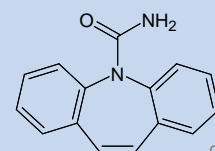
1. Compounds Used in this Study:



Dehydronifedipine

Monoisotopic Mass = 344.100836 Da

MS/MS Transition: 345.1 \rightarrow 284.2



Carbamazepine

Monoisotopic Mass = 236.094963 Da

MS/MS Transition: 237.18 \rightarrow 194.2

2. LC-MS-MS Conditions:

column: Ascentis Express C18, 2 cm x 2.1 mm I.D., 2.7 μm particles (53799-U)
 Discovery C18, 5 cm x 2.1 mm I.D., 5 μm particles (577507-U)
 mobile phase A: 0.1% acetic acid in 95:5 water:acetonitrile
 mobile phase B: 0.1% acetic acid in acetonitrile
 flow rate: see gradient table below
 temp.: 40 $^{\circ}\text{C}$
 det.: ABI 3200 Q Trap MS-MS
 injection: 2 μL
 sample: carbamazepine, dehydronifedipine
 back pressure: both less than 3000 psi

3. Gradient Tables:

column: Ascentis Express, 2 cm x 2.1 mm, 2.7 μm
 flow rate: 1 mL/min

Min	%A	%B
0.00	70	30
0.75	5	95
0.85	5	95
0.90	70	30
1.00	70	30

column: Discovery C18, 5 cm x 2.1 mm, 5 μm
 flow rate: 0.5 mL/min

Min	%A	%B
0.0	70	30
1.5	5	95
1.7	5	95
1.8	70	30
2.0	70	30

Figure 1. Chromatographic Comparison of Fused-Core vs. Porous Particles

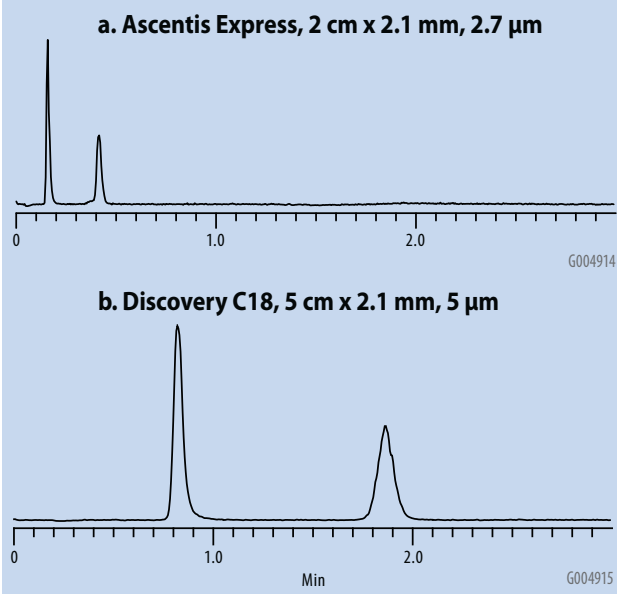


Table 3. Peak Width Comparison

Column	Flow Rate	Peak Width (carbamazepine)	Peak Width (dehydronifedipine)
Ascentis Express C18, 2 cm x 2.1 mm, 2.7 μ m	1 mL/min	0.019	0.028
Discovery C18, 5 cm x 2.1 mm, 5 μ m	0.5 mL/min	0.049	0.095

Figure 2. Calibration Curves

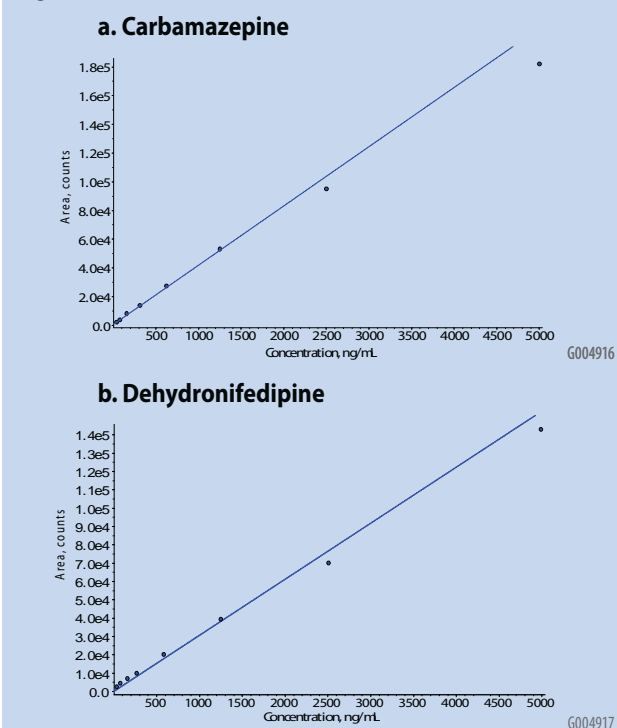


Figure 2 shows a calibration curve for the dehydronifedipine and carbamazepine. The calibration standards were prepared in plasma and extracted using the method in Table 1. Good linearity was obtained over the range studied. Calibration curves are shown using a linear plot with $1/x^2$ weighting.

The demands of increased speed and resolution are vital factors to scientists that work in the area of bioanalytical research. In this work presented here, the benefits of short Fused-Core columns have been demonstrated. The integrity and quality of the analysis was maintained in transferring from totally porous to Fused-Core particles, fast gradients were possible using traditional LC pumps and the Ascentis Express columns have been shown to be rugged and provide good results with greater than 2 years of use (1) in these types of bioanalytical assays.

Reference

- Recent Advancements in Accelerated Bioanalytical LC/MS Using Fused-Core Columns. Richard L. Beardsley, Ethan R. Badman, Zhenmin Liang, Surendra Bansal ASMS Presentation 2009. Philadelphia, PA.

Did you know...?

Supelco's HybridSPE-PPT sample prep cartridges and 96-well plates for the removal of phospholipids are the perfect complement to Ascentis Express for bioanalytical LC-MS-MS assays. If you have an interest in these products, please complete the survey at sigma-aldrich.com/bioanalysis-request

+ Featured Products

Particle Size (μ m)	I.D. (mm)	Length (cm)	Cat. No.
Ascentis Express C18 Columns			
2.7	2.1	2	53799-U
2.7	2.1	3	53802-U
2.7	2.1	5	53822-U
2.7	2.1	7.5	53804-U
2.7	2.1	10	53823-U

+ Related Product

Particle Size (μ m)	I.D. (mm)	Length (cm)	Cat. No.
Discovery C18 Column			
2.7	2.1	5	53799-U

ordering: 800-247-6628 (US only) / 814-359-3441 technical service: 800-359-3041 (US and Canada only) / 814-359-3041

Increased Bioanalytical Throughput and Recovery Utilizing HybridSPE-PPT Small Volume Plates

Craig Aurand

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Often a major concern in developing bioanalytical methods is addressing the affect of biofluid matrix on the detection of desired analytes. The impact of matrix affects in bioanalysis has been well documented. In the majority of cases, co-extracted interferences directly affect the quantitation of analytes due to ionization effects induced by the extracted matrix. This extracted matrix can impact the chromatographic analysis, but more often results in a chromatographic build-up that leads to irregularities in both retention and quantitation. To address these issues, organic gradient elution is often utilized to 'wash' adsorbed contaminants from the column. In most cases gradient elution is not required for resolution of desired analytes, but instead required only to elute extracted matrix from the analytical column.

Performing a more thorough sample cleanup enables faster chromatographic analysis and thus increases the overall sample throughput. Using the HybridSPE-PPT platform for selective phospholipid depletion eliminates the need for gradient elution of adsorbed matrix from the analytical column, resulting in the ability to perform isocratic chromatographic separation with a dramatic increase in throughput.

HybridSPE-Small Volume 96-Well Plate and Closeup of Tips



E001087

Figure 1. HybridSPE-Small Volume 96-Well Schematic Diagram

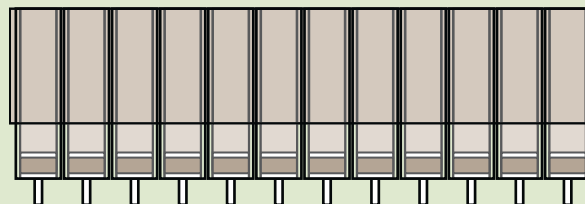
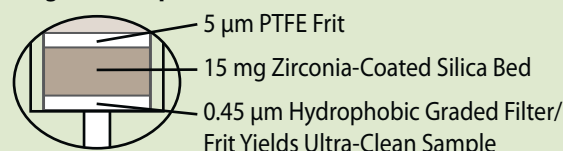


Plate: 96-well format employs special frits at the top and bottom of the same selective bed; proteins **can be removed on-line** for added speed and convenience.

Single Well Explosion:



This study evaluates the performance of the newly developed HybridSPE-PPT Small Volume 96-well plate for preparation of small volumes of rat plasma. The HybridSPE-PPT Small Volume plate accommodates plasma volumes of 20-40 μL , ideally suited for bioanalytical testing of mouse plasma. It makes use of the zirconia-coated silica stationary phase as is also used in the standard HybridSPE-PPT plate for phospholipid depletion. The HybridSPE-PPT Small Volume plate is a scaled down version with a 1 mL well volume and a 15 mg packed stationary bed. A 0.45 μm polishing filter is also used for fine particle removal. The narrow internal well diameter of the plate, along with small packed bed, results in minimal holdup volume (typically 20-40 μL). This enables sufficient volume recovery when handling small volume plasma samples.

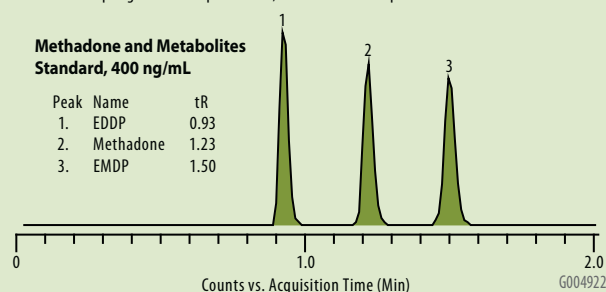
In this study, rat plasma samples spiked with methadone and metabolites 2-ethylidene-1,5-dimethyl-3,3-diphenylpyrrolidine (EDDP) and 2-ethyl-5-methyl-3,3-diphenylpyrrolidine (EMDP) were processed with the HybridSPE-PPT Small Volume 96-well plate and compared against standard protein precipitation methods. The analysis was conducted on an Agilent® 1200SL Rapid Resolution system coupled to an Agilent 6210 TOF LC/MS. Chromatographic separation was performed on the Ascentis Express RP-Amide. The high sensitivity of methadone and metabolites enable for direct small volume injection of the processed sample without the need for evaporation or reconstitution.

Figure 2. Chromatographic Conditions and Conditions for Sample Preparation

column: Ascentis Express RP-Amide, 10 cm x 2.1 mm I.D., 2.7 μ m, (53913-U)
 mobile phase: 10mM ammonium formate (65:35 water:acetonitrile) pH 3.6
 flow rate: 0.4 mL/min
 system pressure: 275 bar
 temp: 35 °C
 inj vol: 0.5 μ L
 instrument: Agilent 1200SL Rapid Resolution, Agilent 6210 TOF LC/MS
 detection: ESI+
 sampling rate: 1.5 spectra/sec, 6309 transients/spectra

Methadone and Metabolites Standard, 400 ng/mL

Peak	Name	tR
1.	EDDP	0.93
2.	Methadone	1.23
3.	EMDP	1.50



Sample Preparation

Standard Solutions:

Standard solutions were prepared from a stock standard in (3:1) 1% formic acid acetonitrile: water at a level of 25, 50, 100, 200, 300 ng/mL. These standards were used to establish recovery of methadone and metabolites from the HybridSPE-Small Volume without interference from plasma interactions

Plasma:

Rat plasma stabilized with K₂EDTA was acquired from Lampire Biological Laboratories, (Pipersville PA). Plasma was spiked directly from stock standard to a level of 100, 200, 400, 800, 1200 ng/mL

HybridSPE-small volume Plasma Samples: apply 20 μ L of plasma to plate, followed by 60 μ L of 1% formic acid acetonitrile. Agitate via vortex for 1 minute, place on vacuum manifold and apply 10" Hg vacuum for 2 minutes. Collect filtrate and analyze directly

HybridSPE-small volume Standard Solution: apply 80 μ L of standard prepared in (3:1) 1% formic acid acetonitrile:water. Agitate via vortex for 1 minute, place on vacuum manifold and apply 10" Hg vacuum for 2 minutes. Collect filtrate and analyze directly. Samples were prepared n=8

Standard Protein Precipitation: apply 100 μ L of plasma to centrifuge vial, followed by 300 μ L of 1% formic acid acetonitrile. Agitate via vortex for 1 minute, place into centrifuge for 2 minutes at 15000 rpm. Collect supernatant and analyze directly

Each spiked level sample was prepared n=8 for both the HybridSPE-PPT Small Volume technique and the standard protein precipitation method. Samples processed using the HybridSPE-PPT Small Volume technique were collected directly into an Agilent low volume 96-well collection plate, average sample volume recovery from the plate was 40 μ L. To ensure that sufficient sample was drawn into the injector, the autosampler was set for bottom well sensing. Samples were assayed for content of methadone and metabolites along with matrix monitoring for phospholipids. In this particular study, monitoring for 1-palmitoylglycerophosphatidylcholine, m/z 496.3375, was conducted as a representative phospholipids matrix ion.

Samples prepared using the HybridSPE-PPT Small Volume plate demonstrated high recovery across the concentration range. These samples were not affected by the matrix buildup due to the complete depletion of phospholipids. No signal suppression was observed using the HybridSPE-PPT Small Volume technique. As shown in Tables 1, 2, and 3 nearly equivalent calibration slopes between the standard solution

Table 1. Calibration Table: Methadone

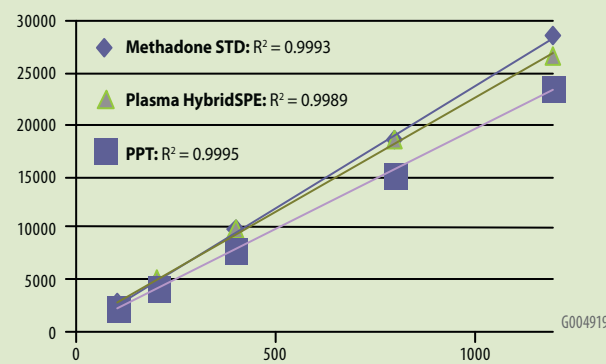


Table 2. Calibration Table: EMDP

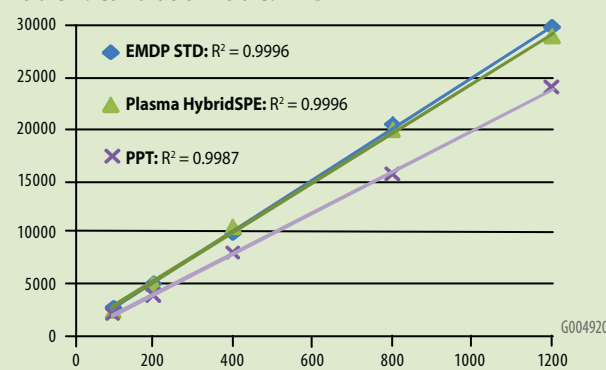
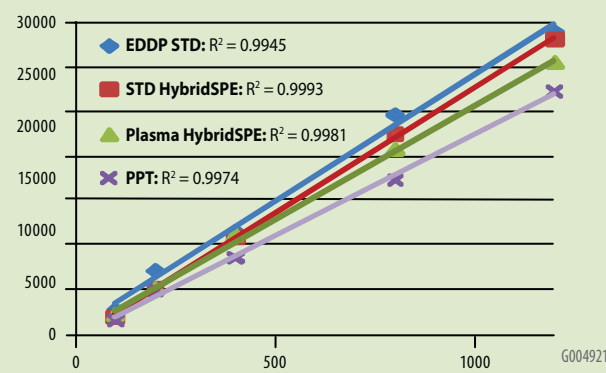


Table 3. Calibration Table: EDDP

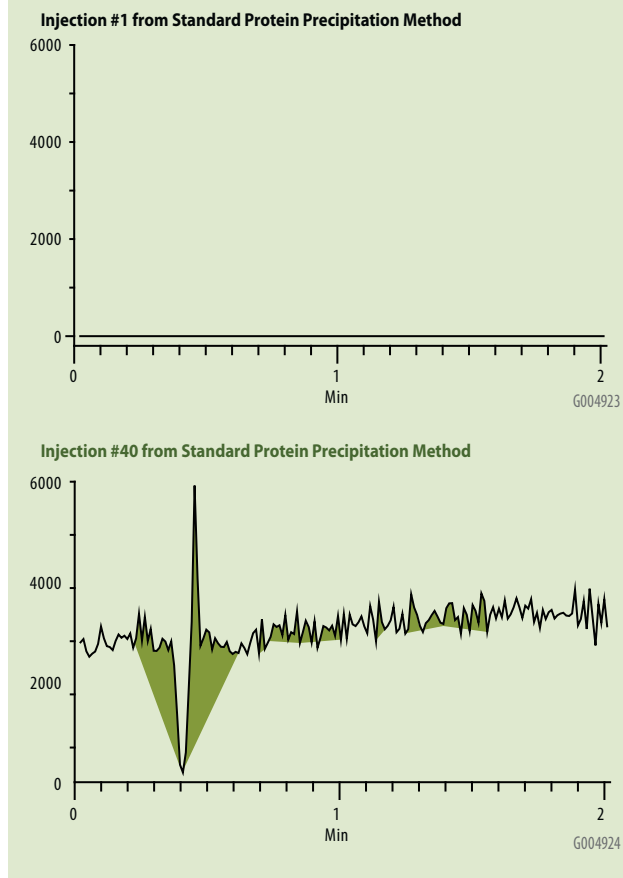


and spiked plasma processed with the HybridSPE-PPT small volume technique was achieved. In Table 3, some drug protein binding was observed for the EDDP metabolite, resulting in the slight decrease in response. The recovery of the standard solution (red) was included in this chart to show the high recovery of EDDP from the HybridSPE-PPT Small Volume plate.

Phospholipid matrix affect was evident with samples prepared using the standard protein precipitation technique. Often when performing ballistic gradient methods, the high organic content elutes a portion of the matrix from the column in a broad range. When performing isocratic

(continued on page 14)

Figure 3. Phospholipid Buildup on Column from Standard Protein Precipitation Method



(continued from page 13)

methods, phospholipid buildup is continuous and results in an overall background increase, due to phospholipids gradually leaching from the column. As demonstrated in Figure 3, monitoring of m/z 496.3375 representing 1-palmitoylglycerophosphatidylcholine, an increase in background from none detected to over 3000 count was observed over the 40 sample injection range of the standard protein precipitation technique. The gradual increase in background phospholipids is the cause for the dramatic decreased signal response with increasing spike level samples. Samples were analyzed in order from lowest spike level to highest spike level. The highest level spiked samples were then subject to the highest amount of chromatographic buildup of phospholipid. Significant signal reduction was observed for the standard protein precipitation method due to the background phospholipids.

This study has demonstrated the detrimental effect of phospholipid buildup and resulting matrix ionization effect when performing standard protein precipitation techniques. By utilizing the HybridSPE-PPT Small Volume plate, excellent recovery of methadone and associated metabolites across the concentration range along with depletion of proteins and phospholipids from the plasma samples was achieved. The combination of facile protein precipitation/phospholipid depletion and fast analysis using modern chromatographic particles shows great promise in increasing the throughput for bioanalytical methods. The ability to perform selective matrix removal enables the use of optimized chromatographic elution conditions without the need for gradient elution of sample matrix, resulting in shorter run times and more rugged bioanalytical methods.

+ Featured Products

Description	Cat. No.
HybridSPE-PPT – Small Volume 96-well plate, 15 mg/well	52794-U
96-Square/Deep Well Collection Plates, 1 mL, PP	575652-U
IKA® MS 3 Digital Orbital Shaker	Z645036-1EA

+ Related Products

Description	Cat. No.
Ascentis Express RP-Amide, 10 cm x 2.1 mm I.D., 2.7 μ m	53913-U
HybridSPE-Precipitation 96-well Plate, 50 mg/well	575656-U
HybridSPE-Precipitation Cartridge, 30 mg/1 mL	55261-U
96-well Protein Precipitation Filter Plate	55263-U
Supelco PlatePrep Vacuum Manifold	57192-U
96-Square/Deep Well Collection Plates, 0.35 mL, PP	575651-U
96-Square/Deep Well Collection Plates, 2 mL, PP	575653-U
96-Square Well Pierceable Cap Mats	575655-U
Methadone, 50 mg and 1 g	M0267
EDDP, 10 mg	E5264

+ Related Information

For more information on the HybridSPE-PPT products, request brochure T409095 (LOP) - *HybridSPE-Precipitation Technology*.

Did you know...?

Supelco's Ascentis Express is a perfect complement to HybridSPE™-PPT sample prep cartridges and 96-well plates for bioanalytical LC-MS-MS assays. If you have an interest in these products, please complete the survey at sigma-aldrich.com/bioanalysis-request.

Introducing Supelco Glass-Fritted Thermal Desorption Tubes with Barcode

Kristen Schultz and Jamie Brown
kristen.schultz@sial.com



E001084

Supelco now offers glass-fritted thermal desorption tubes in the dimension ¼" (6.35 mm) O.D. x 3.5" (89 mm) length for DANI, Markes, OI Analytical, and Shimadzu Instruments.

These tubes are specifically designed to fit any thermal desorber accepting these tube dimensions. In addition to the glass frit at the inlet, each tube is preconditioned, sealed in a TDS³ storage container, and supplied with a unique code-128 barcode. The barcode can easily be read through the TDS³ storage container. The final product is QA tested and packaged in a new field friendly box.

It is a well-documented problem in thermal desorption that glass wool plugs at the inlet have a tendency to blow out of the tube and into the instrument, resulting in sample loss, lost instrument time, contamination of the thermal desorber and instrument repair. Glass frits fused at the optimum position at the inlet prevent this problem from occurring. Other advantages of the glass frit are:

- Optimized position at the inlet maintains the adsorbent(s) in the instruments heated zone. (More consistent recovery of higher molecular weight compounds.)
- Consistent backpressure from tube-to-tube.
- Adsorbent(s) bed(s) stay intact, providing longer serviceable life of the tubes and prevents channeling during sampling.

Note: Over multiple uses, the glass-frit will darken but this does not affect the performance of the tube.

A new feature is the addition of a unique number code-128 barcode, which is easily scanned with any standard barcode reader. If you do not have a barcode reader, the unique number is located directly underneath the barcode and can be manually entered into your system. Having the barcode standard on every tube streamlines sample processing from tube receipt/inventory to deployment and sampling in the field and lastly, to the final steps of sample processing-sample analysis and reporting. It also reduces errors due to unreadable tube markings. Our barcode is Laboratory

Information Management System (LIMS) or like systems friendly and can be integrated into your workflow like other products you currently process with a barcode.

All tubes are preconditioned and QA tested to ensure conformance with our high-quality standards. As the world leader in carbon adsorbent technology and over 20 years of expertise in thermal desorption tube manufacturing and applications, our goal is to ensure you receive the highest quality product.

Each tube is sealed in our TDS³ storage container upon thermal conditioning to preserve the conditioned integrity of the adsorbents. The TDS³ system is unique to Supelco and is an alternative to brass Swagelok® end-fittings, which require tooling for both sealing and removal. The TDS³ storage container does not require any tooling, eliminates internal dead volume, minimizes the risk of contamination from outside sources, and protects the tubes from damage.

Another benefit we often hear from our customers is that the tubes are lighter in weight without the Swagelok® end-fittings and are easier to transport because they are not weighted at the ends. The TDS³ Storage Container System is well-tested in the market with over 15 years of use in various other Supelco thermal desorption tube products such as GERSTEL and Dynatherm and are available for all TD tube sizes.



E001088

+ Featured Products

Description	Qty.	Cat. No.
Tenax® TA Fritted 89 mm Glass TD Tube, w/TDS ³	10	29530-U
Carbopack™ B Fritted 89 mm Glass TD Tube, w/TDS ³	10	29535-U
Carbopack X Fritted 89 mm Glass TD Tube, w/TDS ³	10	29537-U
Carboxen™ 569 Fritted 89 mm Glass TD Tube, w/TDS ³	10	29534-U
Carboxen 1016 Fritted 89 mm Glass TD Tube, w/TDS ³	10	29536-U
Carbotrap™ 217 Fritted 89 mm Glass TD Tube, w/TDS ³	10	29531-U
Carbotrap 300 Fritted 89 mm Glass TD Tube, w/TDS ³	10	29532-U
Carbotrap 349 Fritted 89 mm Glass TD Tube, w/TDS ³	10	29533-U
Empty Fritted 89 mm Glass TD Tube	10	29538-U

ordering: 800-247-6628 (US only) / 814-359-3441 technical service: 800-359-3041 (US and Canada only) / 814-359-3041

Sample Handling

SUPELCO
Analytical

SPME Fast Fit Assemblies (FFA) & Multi-Fiber Exchanger (MFX)

Daniel Vitkuske and Kaj Peterson¹

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1. GERSTEL GmbH & Co.KG, Eberhard-Gerstel-Platz 1, D-45473 Mülheim an der Ruhr

The success of SPME over the past two decades can be attributed to a long list of reasons. Some key points are listed here: SPME is easily automated; a wide range of high-quality fibers are available covering a broad selectivity range; SPME covers analytes ranging from VOCs to SVOCs; additional selectivity in terms of volatility range and matrix interference elimination can be applied by extracting either from the headspace or directly from the liquid sample phase (which can easily be modified to release the maximum amount of VOCs); and last, but not least, the extraction is solvent-free, eliminating dilution and offering a substantial concentration effect along with a reduction in solvent background in the laboratory air.

One thing that many users have sought for quite some time has been the ability to automatically change SPME fibers during the analysis sequence. New SPME Fast Fit Assemblies and the Multi-Fiber Exchanger for the GERSTEL® MultiPurpose Sampler (MPS) now make this possible, adding extra power to SPME automation.

Why Would Anyone Want or Need Automation Power?

There are many benefits to having the ability to automatically exchange SPME fibers. For example, SPME analysis can be automatically performed using fibers with different polarities, covering a wider range of analytes and extracting

Figure 1. Fast Fit Assembly versus a Standard SPME Fiber

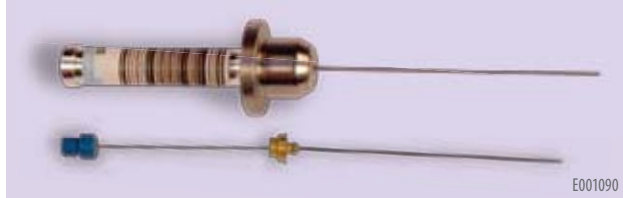


Figure 2. FFA Barcode Guide

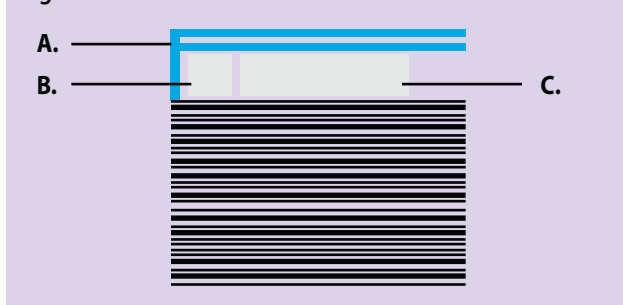


Figure 3. Multi-Fiber Exchanger on the GERSTEL MPS



the maximum amount of information from a sample without manual intervention. MFX enables faster method development by allowing the analyst to modify both method parameters and fiber polarity during an automated sequence. Routine analysis of large batches of difficult samples or samples with a high matrix load can now be performed overnight or through the weekend since fibers can be replaced at regular intervals as the fiber performance deteriorates.

The key to automating fiber exchange is the new SPME Fast Fit Assemblies, which are modified SPME fiber assemblies that allow the MFX system to pickup and operate the fiber assembly without manual intervention. Eliminating the threaded hub and adding certain features (Figure 1) allows the fiber exchanger to readily pick up, expose and retract the fiber, and exchange it with other fiber assemblies.

Special FFA barcoding (Figure 2) provides the user and the autosampler with a variety of information about the properties of the fiber assembly:

- A. The color code indicates the phase type. The coding is the same for the colored screw hub of the traditional SPME fiber assemblies
- B. The first 2 digits of the barcode is the fiber type identity incorporating the phase type & the needle gauge size.
- C. The remaining digits represent a unique tracking number for sample traceability.

The MFX system is available with a three-fiber tray or a 25-fiber tray, in which conditioned SPME FFA's are kept in sealed compartments ready for use. Well-conditioned SPME fibers provide the best possible analytical results and lowest limits of detection. A bake-out station or the GC inlet can be specified for conditioning. If derivatization is needed, pre- or post-extraction derivatization can also be specified.

Stirred, Not Shaken, Sample Extraction

Sample agitation accelerates the SPME process and improves overall throughput, but agitation can lead to mechanical stress on the fiber causing breakage. Sample stirring in the MPS 007 Agitator/Stirrer is based on magnetic stir bars, which do not cause fiber stress and therefore lead to longer fiber life expectancy and improved instrument up-time. The PrepAhead function of the MAESTRO software allows samples to be prepared in advance while the GC run is ongoing for improved throughput and system utilization. In addition, samples can be heated or cooled during storage and extraction, which is highly useful, for example, for fresh food samples.

Summary

The new SPME fast fit assemblies and Multi-Fiber Exchanger were developed in partnership with Chromline srl in Prato, Italy. They offer a unique automated fiber exchange option for the GERSTEL MPS. Currently installed MPS systems, with the MPS L model as the only exception, can be updated to MFX productivity.

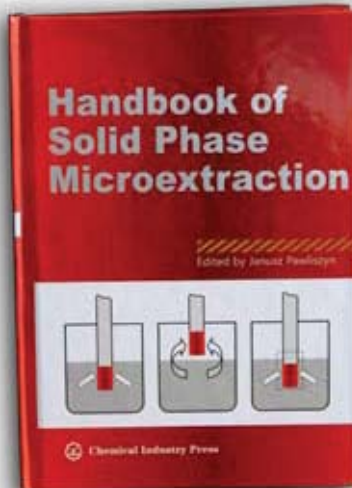
For more information about MFX and other GERSTEL solutions, please contact GERSTEL: gerstel@gerstel.com or visit their website: www.gerstel.com

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SPME FFA Assortment Kits

StableFlex™ Kit, 23 ga 65 µm PDMS/DVB, 50/30 µm DVB/Carboxen/PDMS, 85 µm Carboxen/PDMS, and 85 µm polyacrylate coated fibers	4	FFA57284-U
Kit 1 (Fused silica), 23 ga For Volatiles and Semi-volatiles: 85 µm polyacrylate, 100 µm PDMS, and 7 µm PDMS coated fibers.	3	FFA57285-U
Kit 2 (Fused silica), 23 ga For Volatile or Polar Organics in Water: 75 µm Carboxen/PDMS, 65 µm PDMS/DVB, and 85 µm polyacrylate coated fibers.	3	FFA57286-U
Kit 4 (Fused silica), 23 ga For Flavors and Odors: 100 µm PDMS, 65 µm PDMS/DVB, and 75 µm Carboxen/PDMS coated fibers.	3	FFA57287-U
Kit 5 (Fused silica), 23 ga For Flavors and Odors: 100 µm PDMS, 65 µm PDMS/DVB, 85 µm Carboxen/PDMS, and 50/30 µm DVB/PDMS coated fibers.	4	FFA57362-U

NEW! Handbook of SPME by Janusz Pawliszyn



This new 400-page book contains comprehensive descriptions of the fundamental principles of solid phase microextraction (SPME), recent applications, SPME devices and procedures published to date. SPME protocols are presented in a step-by-step fashion,

providing useful tips and potential pitfalls. The important steps in SPME method development and optimization including calibration are clearly discussed to assist new users of the technology. This handbook enables researchers at all stages of their careers to effectively apply this convenient and solvent free sample preparation technique to solve their analytical challenges in an effective way. This up-to date handbook contains 13 chapters with topics including: Theory of SPME, SPME devices and fiber coatings, commercial devices and coatings, automated SPME systems, calibration of extraction step, SPME method development, ligand-receptor binding, in-vivo SPME, review of different application areas including: environmental, food and fragrance, forensic and drug analysis as well as SPME protocols.

Description	Cat. No.
Handbook of SPME	Z569046

+ Featured Products

Description	Qty.	Cat. No.
SPME Fast Fit Assembly		
100 µm PDMS (Fused silica), 23 ga	3	FFA57341-U
100 µm PDMS (Fused silica), 24 ga	3	FFA57301
30 µm PDMS (Fused silica), 23 ga	3	FFA57289-U
30 µm PDMS (Fused silica), 24 ga	3	FFA57309
7 µm PDMS (Fused silica), 23 ga	3	FFA57291-U
7 µm PDMS (Fused silica), 24 ga	3	FFA57302
85 µm Polyacrylate (Fused silica), 23 ga	3	FFA57294-U
85 µm Polyacrylate (Fused silica), 24 ga	3	FFA57305
60 µm PEG (Metal), 23 ga	3	FFA57354-U
65 µm PDMS-DVB, 23 ga	3	FFA57293-U
65 µm PDMS-DVB, 24 ga	3	FFA57327-U
85 µm CAR/PDMS, 23 ga	3	FFA57295-U
85 µm CAR/PDMS, 24 ga	3	FFA57335-U
50/30 µm DVB/CAR/PDMS, 23 ga	3	FFA57298-U
50/30 µm DVB/CAR/PDMS, 24 ga	3	FFA57329-U

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Sample Handling

SUPELCO
Analytical

FAME Standard for Optimizing GC System Performance

Vicki Yearick and Katherine Stenerson

vicki.yearick@sial.com

AOCS and AOAC methods for determining the trans fatty acid composition of foods by gas chromatography require the use of a highly polar cyanosilicone capillary column to provide the best peak resolution attainable of the numerous geometric (cis & trans) and positional isomers. Due to the highly polar nature of these phases, subtle changes in the GC system can alter the chromatographic separation of the positional cis & trans isomers. Therefore, it is important to continually monitor column performance and correct for unexpected changes in the column flow, oven temperature, column degradation and/or sample concentration as they occur.

AOCS Method Ce 1h-05 recommends using a well-characterized mixture of fatty acid methyl esters (FAMES) covering the range of fatty acids under investigation to monitor changes in column performance over time. Sigma-Aldrich's new cis/trans FAME Column Performance Mix is an excellent choice for monitoring these changes. This qualitative mix is specifically designed to optimize and to monitor performance changes per AOCS Method Ce 1h-05 and AOAC Method AOAC 996.06. The mix contains C18:1 cis/trans positional isomers of critical importance to the food industry.

Because some cis/trans positional FAMES are difficult to separate and baseline resolution may not be possible, regular use of the Sigma-Aldrich FAME Column Performance Mix is of utmost importance to achieve the required column performance. Small changes in the sample size, sample concentration, or oven temperature may be required to achieve the best resolution between the C18:1 cis/trans isomers. Once optimal resolution for the column is established, weekly analysis of the mix allows monitoring of the column's performance.

The analyst injects the mix and identifies the peaks labeled as "1" and "2" in Figure 1. The resolution between these peaks is then calculated using the equation shown in Figure 2. It is important to run the mix upon initial installation of a new capillary column to ensure that conditions are established to optimize resolution. Optimum resolution between two peaks is defined as "baseline" resolution. Full baseline resolution is achieved when the R-value calculated using the equation in Figure 2 is approximately 1.5 or greater. The example chromatogram shown in Figure 1 illustrates the measurements required when calculating the resolution value - specifically peak width and retention time. As is evident in Figure 1, baseline resolution of peaks 1 and 2 will not be attained under normal testing conditions. The minimum acceptable resolution value will be described in a future revision of AOCS Method Ce 1h-05. For now, laboratories must establish their own criteria.

Figure 1. cis/trans FAME Column Performance Mix

column: SP™-2560, 100 m x 0.25 mm I.D., 0.20 µm (24056)
 oven: 180 °C (isothermal)
 inj: 250 °C
 det.: FID, 250 °C
 carrier gas: helium, 1 mL/min. @ 180 °C
 injection: 1 µL, 100:1 split
 sample: cis/trans FAME column performance mix (40495-U)

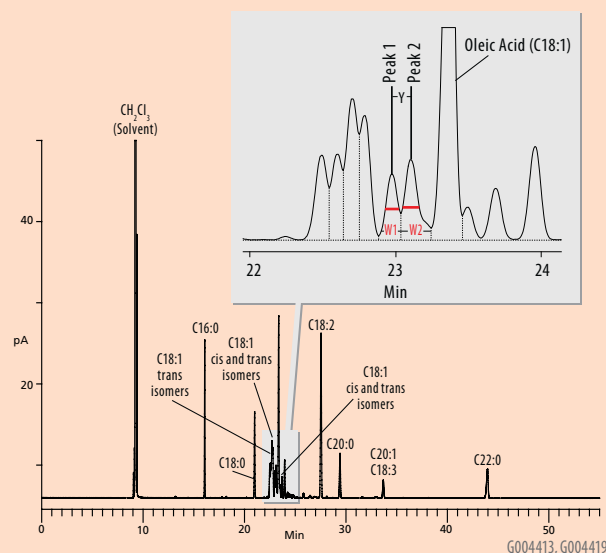


Figure 2. Resolution Equation

$$R = 1.18 (t_{R2} - t_{R1}) / (w_{(1/2)1} + w_{(1/2)2})$$

t_{R1} = the retention time of peak 1

t_{R2} = the retention time of peak 2

$w_{(1/2)1}$ = peak width at half height of peak 1 in mm

$w_{(1/2)2}$ = peak width at half height of peak 2 in mm

For example:

$$R = 1.18(23.053 - 22.922) / (0.0846 + 0.0912)$$

$$R = 0.88$$

At present, no column will fully resolve all C18:1 cis/trans positional isomers. By utilizing the cis/trans FAME Column Performance Mix, analysts can optimize column performance before assaying complex mixtures. Regular use also makes it possible to monitor column degradation and to recognize when to replace the capillary column.

References

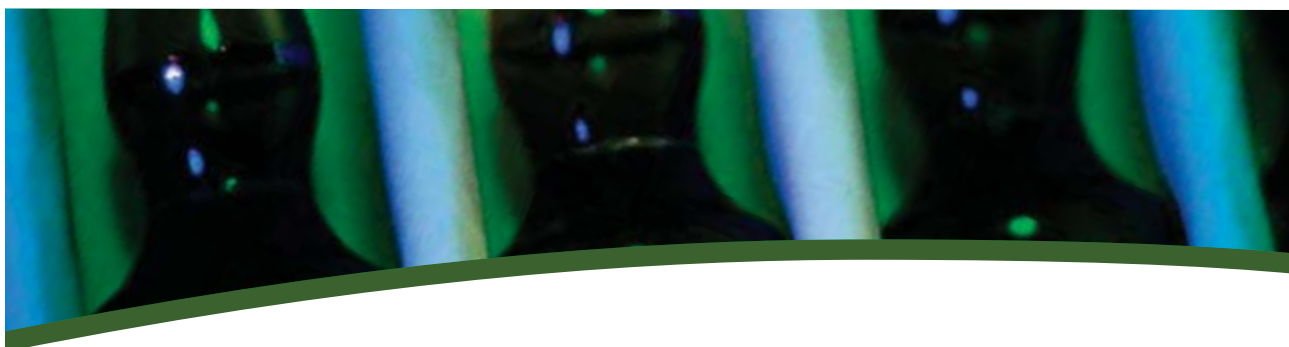
1. AOCS Official Method Ce 1h-05. Determination of cis-, trans-, Saturated Monosaturated and Polysaturated Fatty Acids in vegetable or Non-ruminant Animal Oils and fats by Capillary GLC, 2005.
2. AOAC Official Method Ce 996.06. Fat (Total, Saturated, and Unsaturated) in Food, Hydrolytic Extraction Gas Chromatographic Method, 2001.

+ Featured Products

Description	Cat. No.
cis/trans FAME Column Performance Mix, 2.5 mg/mL	
1 x 1mL	40495-U
10 x 1mL	4M0495-U

+ Related Products

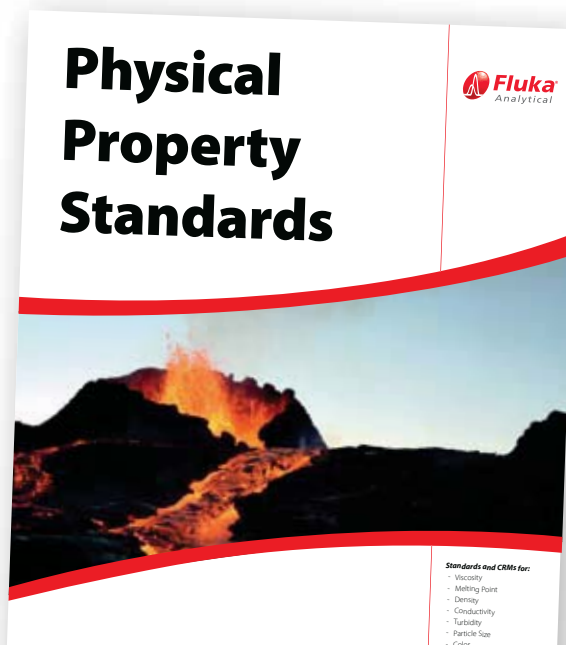
Description	Cat. No.
SP-2560 Column	
100 m x 0.25 mm I.D., 0.20 μ m	24056
75 m x 0.18 mm I.D., 0.14 μ m	23348-U



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Molded Thermogreen LB-2 Septa Durability

Robert F. Wallace and Gregory A. Baney
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Introduction

The injection port septa selected for gas chromatography (GC) use should exhibit low bleed, resist leaks, and be easy to penetrate. Molded Thermogreen LB-2 septa have all these characteristics. Using the same exclusive rubber formulation as the traditional Thermogreen LB-2, molded Thermogreen LB-2 septa allow for stricter tolerances of both diameter and thickness, resulting in much easier installation and better sealing. A previous article discussed the low bleed characteristics of molded Thermogreen LB-2 septa (1). This article will focus on their resistance to leaks.

Durability Requirements

A GC septum should resist fragmentation. This is because septum fragments in the inlet liner bleed (observed as extraneous peaks) and adsorb active sample components (causing loss of analyte response). Molded Thermogreen LB-2 septa are specially engineered to offer excellent resistance to fragmentation. With its high elasticity, molded Thermogreen LB-2 septa ensure that multiple injections can be obtained

with a wide range of needle compatibility. In fact, the entire Thermogreen LB-2 family is widely accepted as the industry standard in GC septa because of their great durability.

Puncturability Test

As a measure of durability, a molded Thermogreen LB-2 septum with injection hole was subjected to repeated injections. Column head pressure and carrier gas linear velocity/flow rate were monitored throughout the test to determine if the septum developed a leak. An autosampler was used for the test, ensuring that the syringe needle (tapered 23s-26s gauge) punctured the same location repeatedly. Figure 1 shows chromatograms from the initial run and after 2100 injections. As demonstrated, no artifacts were present in either the initial or final runs to indicate any problems with the chromatography. Additionally, retention, head pressure, and column flow remained constant, indicating that a leak-free seal was maintained.

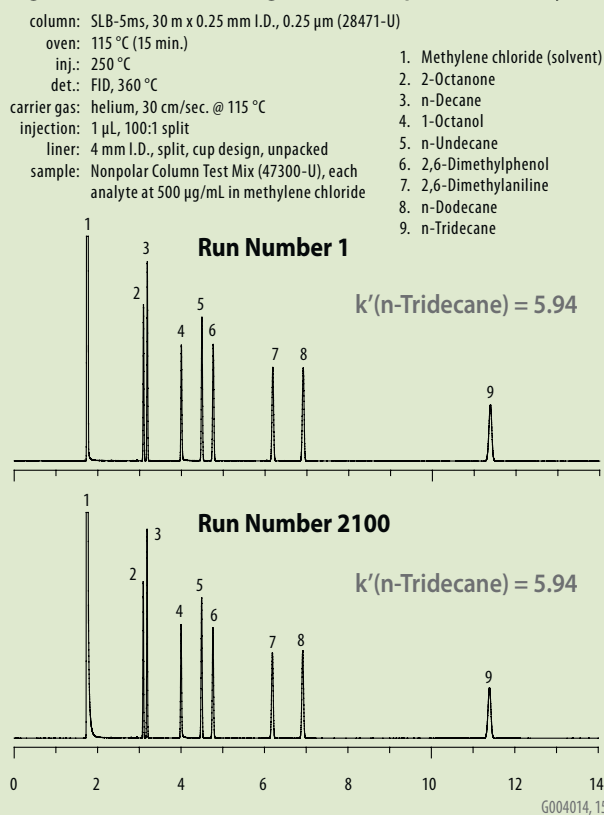
Conclusion

The quality traits to look for in a GC septum are low bleed, resistance to leaks, and easy penetration. The strict tolerances of the mold used to manufacture molded Thermogreen LB-2 septa result in septa that are easier to install and consistently fit better. These septa exhibit an ultra low bleed profile, are temperature stable to 350 °C, are very resistant to both slivering and coring, and have a high puncture tolerance.

Reference

1. M.D. Buchanan, "NEW! Molded Thermogreen LB-2 Septa" The Reporter 25.3 (2007) p. 15.

Figure 1. Molded Thermogreen LB-2 Septum Durability Test



Featured Products

Diam. (mm)	Type	Qty.	Cat. No.
Molded Thermogreen LB-2 Septa			
9.5 (3/8 in.)	with injection hole	50	28331-U
9.5 (3/8 in.)	with injection hole	250	28332-U
9.5 (3/8 in.)	solid disc	50	28670-U
9.5 (3/8 in.)	solid disc	250	28671-U
10 (13/32 in.)	with injection hole	50	28333-U
10 (13/32 in.)	with injection hole	250	28334-U
10 (13/32 in.)	solid disc	50	28673-U
10 (13/32 in.)	solid disc	250	28675-U
11 (7/16 in.)	with injection hole	50	28336-U
11 (7/16 in.)	with injection hole	250	28338-U
11 (7/16 in.)	solid disc	50	28676-U
11 (7/16 in.)	solid disc	250	28678-U

Related Products

Description	Cat. No.
Hook septum puller	20352
Screw septum puller	20353

SGE Introduces Color-Coded Syringes



E001098, E001099, E001100

SGE has implemented significant changes to their chromatographic syringe line to make it easier to distinguish between the different volumes of syringes while experiencing improved technical performance.

The glass barrels of SGE autosampler syringes are now color-coded according to volume to make it easier and faster for an analyst to select the proper volume of syringe for different applications. For example, the 50 μL syringe has a purple barrel while the 10 μL syringe can be distinguished by its orange barrel. This color-coding also makes it easier for quick volume identification of syringes already installed in an autosampler.

The new color-coded syringes are now packaged in a clear 100% recyclable protective inner sleeve with a fully recyclable outer box with a window for easy visual identification of the product when stored in a drawer.



E001101

Newly introduced tighter physical specifications and design changes have greatly reduced or eliminated common problems including syringe adhesive contamination of the sample, sample carryover, and mechanical problems with the needle, plunger and barrel. These improvements provide superior technical performance for the analysts and have the added benefit of extending the life of the syringe.

Longer Life

Modifications to the inner surface finish of the syringe glass barrel and the design of the syringe components provide greater solvent resistance, a wider working-temperature range and improved operational smoothness. As a result of these modifications, the lifetime of the syringe can be as much as ten times greater when compared to SGE's current syringes.

Superior Performance

Elimination of syringe adhesive from the fluid path removes the possibility of the sample being contaminated by adhesive, sample carryover, or the possibility of the sample matrix dissolving the adhesive that holds syringe components together. This is accomplished by the addition of a press fit PTFE seal that fits tightly against the inner walls of the syringe and is flush to the end of the needle leaving no gaps and preventing contact of the sample with the syringe adhesive.

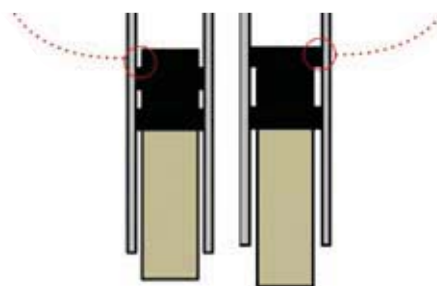
Additionally, the use of a new adhesive chemistry in the manufacturing of SGE syringes allows the syringe to be operated over a wider temperature range.

Improvements have also been made to the needle design. The needle and hub are permanently fused together, increasing the strength of the parts.

Reduced Carryover

Design improvements in the PTFE plunger tips and the PTFE seal inserts mentioned previously create a much tighter fit between the plunger tip of the barrel and the PTFE seal. A smoother glass surface ensures that the fluid is flushed out and no carryover occurs.

Superior Plunger Tip



Other tip designs make it impossible to completely expel sample.

SGE 2010 plunger tip sits flush against syringe insert, minimizing any potential carryover.

SGE has made significant changes to improve their syringe line for 2010. The changes in both packaging and syringe design make these products better with extended life, easier identification, and overall better performance for syringe customers.

For more information on any of our syringes, email our Technical Service Department at techservice@sial.com or visit us on the web at sigma-aldrich.com/syringes.

The Utility of Headspace Grade Solvents for the Analysis of Organic Volatile Impurities

Katherine Stenerson and Shyam Verma

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Introduction

Organic volatile impurities or “OVIs” sometimes remain behind in pharmaceutical preparations as a result of synthetic and manufacturing procedures. For reasons of health and safety, testing is done to ensure that these solvents are not above concentration limits listed by the United States Pharmacopeia (USP) and in the International Conference on Harmonization (ICH) guidelines (1,2). Static headspace GC (SH-GC) is a commonly used technique in the analysis of these OVIs. This technique concentrates volatile analytes, and allows for their analysis free from sample matrix.

Samples to be analyzed by SH-GC must be dissolved in a suitable solvent; and in addition to being able to dissolve the sample, the solvent chosen must allow for sufficient sensitivity of the analytes of interest in the headspace. Partition coefficient affect the ability of an analyte to enter into the headspace from the liquid sample, and those with lower values will have higher vapor pressure and thus greater sensitivity in the headspace (3). Thus, for low-level detection, the analytes of interest must have low partition coefficients in the dissolution solvent chosen. In addition, the vapor pressure of the dissolution solvent itself should be sufficiently low so it will not affect detection of OVI analytes by “flooding” the headspace.

Water offers a very low partition coefficient for analytes and has a low vapor pressure, but cannot be used in all cases. USP <467> and European Pharmacopeia (EP) methodologies list procedures for both water soluble and water-insoluble samples. For water-insoluble samples, USP <467> designates the use of the solvents dimethyl sulfoxide (DMSO) and dimethylformamide (DMF). Other dissolution solvents that have been found to be useful for headspace analysis of water-insoluble samples include dimethylacetamide (DMAC) and 1,3-dimethyl-2-imidazolidinone (DMI), and the later is described for use in EP Method 2.4.24 (4). These solvents elute later than most OVI analytes in chromatographic analyses, and have significantly lower vapor pressure than many other high boiling-point organic compounds.

The purity of dissolution solvents is essential to avoid extraneous peaks in the chromatographic analysis, and prevent interference with the analytes of interest. Many protocols followed by laboratories doing OVI analysis require the analysis of an acceptable blank, and some published methodologies, such as EP Method 2.4.24,

require the analysis of a blank to verify the absence of interfering peaks. Fluka brand high purity headspace grade solvents are now available which are specially tested to ensure suitability for SH-GC analysis.

Experimental

In this evaluation, the utility of Fluka brand DMSO headspace grade solvent was evaluated for use in the SH-GC analysis of OVIs. The purity of the headspace solvent was evaluated by preparing sample blanks using headspace and alternative grades of DMSO. The retention times of peaks present in the blanks were then compared to an OVI standard prepared in headspace grade DMSO solvent.

The blanks were prepared by pipetting 1 mL of DMSO into a 10 mL headspace vial, and subjecting the sealed vial to SH-GC analysis. The OVI working standard included a variety of common process solvents, representing various classes as described in USP <467> and ICH guidelines. It was prepared from a stock solution in DMAC by dilution with headspace grade DMSO. The composition and final concentration of the OVI working standard is summarized in Table 1. The working standard was prepared for SH-GC analysis in a similar manner

Table 1. Composition of OVI Standard + Peak IDs for Figure 3

Peak #	Compound	Class	Concentration (µg/mL)	Cat. No.
1	Methanol	2	237	82762
2	Ethanol	3	395	–
3	Acetone	3	235	90872
4	Isopropanol	3	390	–
5	Acetonitrile	2	39	45983
6	Methylene chloride	2	66	02575
7	tert-Butanol	NC	237	50621
8	Methyl-tert-butyl ether	3	221	08603
9	n-Hexane	2	33	52750
10	n-Propanol	3	240	96566
11	Methyl ethyl ketone	3	240	02469
12	Ethyl acetate	3	271	58958
13	Tetrahydrofuran	2	88	78445
14	Cyclohexane	2	232	–
15	Isobutanol	3	241	82059
16	Isopropyl acetate	3	262	90871
17	2-Methyl-tetrahydrofuran	NC	86	85488
18	n-Heptane	3	68	51730
19	1-Methoxy-2-propanol	NC	192	65692
20	1,4-Dioxane	2	52	76887
21	4-Methyl-2-pentanone	3	239	02474
22	Toluene	2	86	89680
23	Diisopropyl ethylamine	NC	94	–
24	Dimethylformamide	2	74	72438
25	m-Xylene	2	86	95670
26	o-Xylene	2	88	95660
27	Tetramethyl urea	NC	387	–
28	N-Methylpyrrolidone	2	614	78769

NC = solvent not classified

to the blanks. Both blanks and the standard were analyzed by SH-GC using the parameters listed in Table 2.

Results

Chromatograms of the SH-GC blanks using headspace and organic synthesis grades of DMSO are presented in Figures 1 and 2. Overall, the headspace grade blank had fewer peaks in the OVI elution range than the organic synthesis grade blank. Comparing the blanks with the chromatogram of the working OVI standard prepared in headspace-grade DMSO

Table 2. SH-GC Parameters

Headspace Parameters		
Temperatures:	Times:	Pressures:
Oven: 100 °C Loop: 110 °C Transfer Line: 150 °C	Vial Equilibration: 10 min. Pressurization: 0.2 min. Loop Fill: 0.2 min. Loop Equilibration: 0.05 min.	Vial: 15 psi Transfer Line: 25 psi Loop Volume: 1.0 mL Inject Time: 1 min.
GC Parameters		
column: SPB™-624, 30 m x 0.32 mm I.D., 1.8 µm (28464-U)		
oven: 40 °C (4 min.), 8 °C/min. to 60 °C, 5 °C/min. to 85 °C (2 min.), 30 °C/min. to 220 °C (2 min.)		
inj.: 225 °C		
det.: FID, 270 °C (FID)		
carrier gas: Helium, 1.5 mL/min, constant		
injection: 1.0 mL, 5:1 split		
liner: 2 mm I.D.		

Figure 1. SH-GC Blank, DMSO – Organic Synthesis Grade

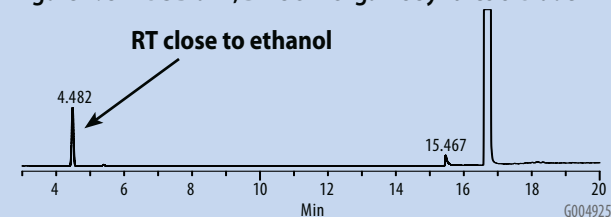


Figure 2. SH-GC Blank, DMSO – Fluka Headspace Grade

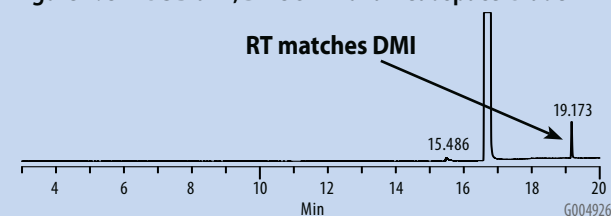
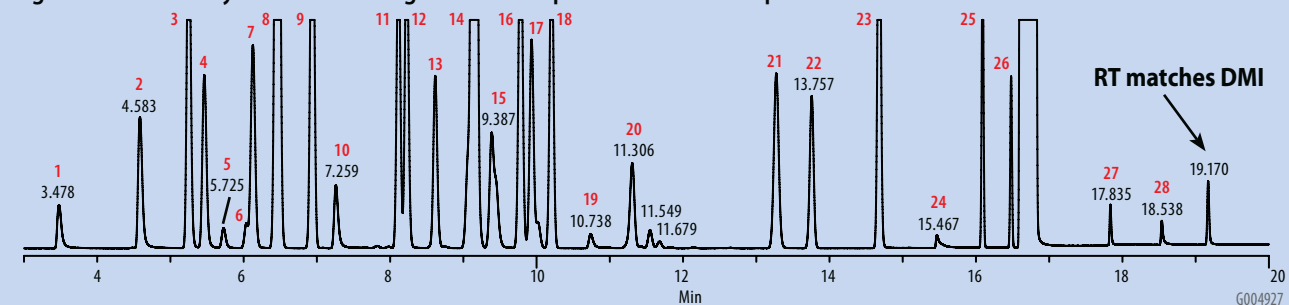


Figure 3. SH-GC Analysis of OVI Working Standard Prepared in Fluka Headspace Grade DMSO



(Figure 3), both blanks were found to contain some DMF. The organic synthesis blank contained a peak eluting close to the retention time of ethanol. This peak could potentially interfere with the proper detection and analysis of ethanol as an OVI. A peak corresponding to the retention time of DMI was detected in the SH-GC blank. This same peak was also detected in the OVI working standard, which was prepared in headspace grade DMSO.

Conclusions

The Fluka headspace grade produced a “cleaner” blank for SH-GC analysis than a lesser grade of DMSO. The peaks that did appear in the headspace grade solvent blank eluted outside of the retention range of the OVIs analyzed. Additional headspace grade solvents (DMF, DMAC, and DMI) suitable for this application are also available. Experimental data on these solvents, similar to that presented here for DMSO, can be obtained by requesting Supelco Publication T409180 (only available electronically).

Acknowledgements

We would like to thank Amanda Quiroga and Michael Dong at Genentech USA, for providing the data for this evaluation, and for their invaluable input.

References

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- ICH Guidelines for Industry, Q3C Impurities: Residual Solvents, US Dept. of Health and Human Services Food and Drug Administration, Center for Drug Evaluation and Research (CDER), Center for Biologics Evaluation and Research (CBER), ICH December, 1997.
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- European Pharmacopoeia (EP) 5.0, Vol.1, (2004), 2.4.24 Identification and Control of Residual Solvents.
- A. Quiroga, M. Dong, K. Stenerson, S. Verma, The Utility of Headspace Grade Solvents in the Analysis of Organic Volatile Impurities. Supelco Publication T409180, November 2009.

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Dimethyl sulfoxide	189 °C	1 L	51779
N,N-Dimethylformamide	153 °C	1 L	51781
1,3-Dimethyl-2-imidazolidinone	225 °C	1 L	67484
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